

PATENT APPLICATION TRANSMITTAL LETTER
(Small Entity)

Docket No.
0109015/016

TO THE ASSISTANT COMMISSIONER FOR PATENTS

Transmitted herewith for filing under 35 U.S.C. 111 and 37 C.F.R. 1.53 is the patent application of:

Renji Yang and Karl K. Johe

For: **STABLE NEURAL STEM CELL LINES**

Enclosed are:

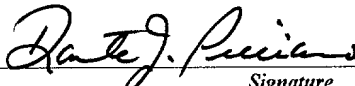
- ☒ Certificate of Mailing with Express Mail Mailing Label No. **EJ091608439US**
- ☒ **20 pages + 11** sheets of drawings.
- ☐ A certified copy of a _____ application.
- ☒ Declaration ☒ Signed. ☐ Unsigned.
- ☒ Power of Attorney
- ☐ Information Disclosure Statement
- ☐ Preliminary Amendment
- ☒ **two** Verified Statement(s) to Establish Small Entity Status Under 37 C.F.R. 1.9 and 1.27.
- ☒ Other: **Cover Sheet and Assignment**

CLAIMS AS FILED

For	#Filed	#Allowed	#Extra	Rate	Fee
Total Claims	22	- 20 =	2	x \$9.00	\$18.00
Indep. Claims	4	- 3 =	1	x \$39.00	\$39.00
Multiple Dependent Claims (check if applicable) <input type="checkbox"/>					\$0.00
BASIC FEE					\$380.00
TOTAL FILING FEE					\$437.00

- ☐ A check in the amount of _____ to cover the filing fee is enclosed.
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Dated: **September 20, 1999**



Dante J. Picciano
Reg. No. 33,543

CC:

**VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY
STATUS (37 CFR 1.9(f) AND 1.27 (b)) - INDEPENDENT INVENTOR**

Docket No.
0109015/016

Serial No.

-

Filing Date

September 20, 1999

Patent No.

-

Issue Date

-

Applicant/ **Renji Yang and Karl K. Johe**
Patentee:

Invention: **STABLE NEURAL STEM CELL LINES**

As a below named inventor, I hereby declare that I qualify as an independent inventor as defined in 37 CFR 1.9(c) for purposes of paying reduced fees under section 41(a) and (b) of Title 35, United States Code, to the Patent and Trademark Office with regard to the invention entitled above and described in:

- ☒ the specification to be filed herewith.
☐ the application identified above.
☐ the patent identified above.

I have not assigned, granted, conveyed or licensed and am under no obligation under contract or law to assign, grant, convey or license, any rights in the invention to any person who could not be classified as an independent inventor under 37 CFR 1.9(c) if that person had made the invention, or to any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).

Each person, concern or organization to which I have assigned, granted, conveyed, or licensed or am under an obligation under contract or law to assign, grant, convey, or license any rights in the invention is listed below:

- ☐ No such person, concern or organization exists.
☒ Each such person, concern or organization is listed below.

*NOTE: Separate verified statements are required from each named person, concern or organization having rights to the invention averring to their status as small entities (37 CFR 1.27)

FULL NAME **NeuralStem Biopharmaceuticals, Ltd.**

ADDRESS **387 Technology Drive, College Park, MD 20742**
☐ Individual ☒ Small Business Concern ☐ Nonprofit Organization

FULL NAME _____

ADDRESS _____

☐ Individual ☐ Small Business Concern ☐ Nonprofit Organization

FULL NAME _____

ADDRESS _____

☐ Individual ☐ Small Business Concern ☐ Nonprofit Organization

FULL NAME _____

ADDRESS _____

☐ Individual ☐ Small Business Concern ☐ Nonprofit Organization

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b))

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

NAME OF INVENTOR Renji Yang

SIGNATURE OF INVENTOR _____

DATE: _____

NAME OF INVENTOR Karl K. Johe

SIGNATURE OF INVENTOR _____

DATE: _____

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NAME OF INVENTOR Renji Yang

SIGNATURE OF INVENTOR Renji Yang

DATE: 9/17/99

NAME OF INVENTOR Karl K. Johe

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**VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY
STATUS (37 CFR 1.9(f) AND 1.27 (c)) - SMALL BUSINESS CONCERN**

Docket No.
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September 20, 1999

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Applicant/ **Renji Yang and Karl K. Johe**
Patentee:

Invention: **STABLE NEURAL STEM CELL LINES**

I hereby declare that I am:

- ☐ the owner of the small business concern identified below:
☒ an official of the small business concern empowered to act on behalf of the concern identified below:

NAME OF CONCERN: **NeuralStem Biopharmaceuticals, Ltd.**

ADDRESS OF CONCERN: **387 Technology Drive, College Park, MD 20742**

I hereby declare that the above-identified small business concern qualifies as a small business concern as defined in 37 CFR 121.3-18, and reproduced in 37 CFR 1.9(d), for purposes of paying reduced fees under Section 41(a) and (b) of Title 35, United States Code, in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third party or parties controls or has the power to control both.

I hereby declare that rights under contract or law have been conveyed to and remain with the small business concern identified above with regard to the above identified invention described in:

- ☒ the specification filed herewith with title as listed above.
☐ the application identified above.
☐ the patent identified above.

If the rights held by the above-identified small business concern are not exclusive, each individual, concern or organization having rights to the invention is listed on the next page and no rights to the invention are held by any person, other than the inventor, who could not qualify as an independent inventor under 37 CFR 1.9(c) or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).

Each person, concern or organization to which I have assigned, granted, conveyed, or licensed or am under an obligation under contract or law to assign, grant, convey, or license any rights in the invention is listed below:

- ☒ no such person, concern or organization exists.
☐ each such person, concern or organization is listed below.

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 ADDRESS _____

☐ Individual ☐ Small Business Concern ☐ Nonprofit Organization

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NAME OF PERSON SIGNING: I. Richard Garr

TITLE OF PERSON SIGNING _____

OTHER THAN OWNER: President and CEO

ADDRESS OF PERSON SIGNING: NeuralStem Biopharmaceuticals, Ltd.

387 Technology Drive

College Park, MD 20742

SIGNATURE: _____ DATE: _____

any rights in the invention is listed below:

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NAME OF PERSON SIGNING:

I. Richard Garr

TITLE OF PERSON SIGNING

OTHER THAN OWNER:

President and CEO

ADDRESS OF PERSON SIGNING:

NeuralStem Biopharmaceuticals, Ltd.387 Technology DriveCollege Park, MD 20742

DATE:

2/17/27

Stable Neural Stem Cell Lines

This patent application is directly related to U.S. provisional patent application 60/101,354, filed September 22, 1998, and is a continuation-in-part of U.S. patent application 09/053,414, filed April 1, 1998, now pending, which is a continuation-in-part of U.S. patent application 08/719,450, filed September 25, 1996, now U.S. Patent 5,753,506, the entire contents of each is hereby incorporated by reference and relied upon.

Background of the Invention

1. FIELD OF THE INVENTION

The present application discloses a systematic and efficient method for establishing stable neural stem cell lines and neuronal progenitor lines. The resulting cell lines provide robust, simple, and reproducible cultures of human and other mammalian neurons in commercially useful mass quantities while maintaining normal karyotypes and normal neuronal phenotypes.

2. DESCRIPTION OF THE RELATED ART

A developing fetal brain contains all of the cells germinal to the cells of an adult brain as well as all of the programs necessary to orchestrate them toward the final network of neurons. At early stages of development, the nervous system is populated by germinal cells from which all other cells, mainly neurons, astrocytes and oligodendrocytes, derive during subsequent stages of development. Clearly such germinal cells that are precursors of the normal brain development would be ideal for all gene-based and cell-based therapies if these germinal cells could be isolated, propagated and differentiated into mature cell types.

The usefulness of the isolated primary cells for both basic research and for therapeutic application depends upon the extent to which the isolated cells resemble those in the brain. Just how many different kinds of neural precursor cells there are in the developing brain is unknown. However, several distinct cell types may exist:

- a unipotential precursor to neurons only ("committed neuronal progenitor" or "neuroblast"),
- a unipotential precursor to oligodendrocytes only ("oligodendroblast"),
- a unipotential precursor to astrocytes only ("astroblast"),
- a bipotential precursor that can become either neurons or oligodendrocytes, neurons or astrocytes, and oligodendrocytes or astrocytes, and

a multipotential precursor that maintains the capacity to differentiate into any one of the three cell types.

CNS stem cells are multipotential precursor cells with the innate property to differentiate into all major cell types of the mammalian central nervous system (CNS) including neurons, astrocytes, and oligodendrocytes. The methods for isolation and differentiation of CNS stem cells and the characterization of differentiated cell types have been previously described in detail, U.S. Patent 5,753,506 (Johe). Briefly, CNS stem cells are expanded in serum-free, chemically defined medium containing basic fibroblast growth factor, bFGF, as the sole mitogen. The culture condition permits nearly pure populations of CNS stem cells for a long period both as a mass culture and as a clonal culture.

The mitotic capacity of CNS stem cells, however, is finite. With the previous culture conditions, it had been difficult to expand CNS stem cells beyond about 30 cell-doublings at which point a majority of the cells have lost their capacity for neuronal differentiation and further expand as glial progenitors rather than as multipotential stem cells. The mechanism for this limitation is yet unknown.

We hypothesized that mitotic CNS stem cells secrete an autocrine factor or factors which suppress the entry into cell cycle at the G1 phase of mitosis. This would effectively antagonize the mitogenic actions of bFGF and initiate the differentiation path. Thus, it is a mechanism to self-regulate the proliferation of CNS stem cells and, *in vivo*, to limit the generation of neurons and glia during development. Consistent with this mechanism is the observation that high cell density promptly differentiates CNS stem cells even in the presence of bFGF and regardless of the passage time.

Although the 30 cell-doublings yield 10^9 -fold expansion of cells, a method for further significant expansion of CNS stem cells would be of significant commercial value. Here, we disclose that constitutive activation of c-myc protein in CNS stem cells prevents their spontaneous differentiation at high cell density, confers resistance to glial differentiation, and increases the mitotic capacity over 60 cell-doublings. This procedure thus yields more than a 10^{18} -fold expansion of CNS stem cells.

SUMMARY OF THE INVENTION

The present application reveals a method for producing stable cell lines of mammalian neural precursor cells *in vitro*. The method comprises the steps of preparing a culture of neural precursor cells in a serum-free medium; culturing the neural precursor cells in the presence of a first mitogen, where the first mitogen is selected from the group consisting of aFGF, bFGF, EGF, TGF α and combinations thereof; contacting the cells with an agent capable of being taken up by the cells and capable of expressing a c-myc gene; and further culturing the cells in a medium containing the first mitogen and a second mitogen, where the second mitogen is selected from the group consisting of aFGF, bFGF, EGF, TGF α , serum and

combinations thereof, with the proviso that the second mitogen is other than the first mitogen.

In a preferred embodiment of the method, the c-myc gene is fused with other DNA elements, where the other DNA elements comprise at least one element selected from the group consisting of a ligand binding domain for an estrogen receptor, an androgen receptor, a progesterone receptor, a glucocorticoid receptor, a thyroid hormone receptor, a retinoid receptor, and an ecdysone receptor.

In another preferred embodiment of the method, the medium containing the first mitogen and the second mitogen further comprises a myc-activating chemical selected from the group consisting of β -estradiol, RU38486, dexamethasone, thyroid hormones, retinoids, and ecdysone.

In a more preferred embodiment of the method, the mammalian neural precursor cells are derived from a human. In another more preferred embodiment of the method, the mammalian neural precursor cells are derived from an *in vitro* culture of pluripotent embryonic stem cells.

The present application also reveals a cell line produced according to this method. In a preferred embodiment of the cell line, the cells maintain a multipotential capacity to differentiate into neurons, astrocytes and oligodendrocytes. In other preferred embodiments of the cell line, the cells maintain a bipotential capacity to differentiate into neurons and astrocytes or into astrocytes and oligodendrocytes.

In more preferred embodiments of the cell line, the cells maintain a unipotential capacity to differentiate into neurons or into astrocytes.

The present application also reveals a method for producing stable clonal cell lines of mammalian neural precursor cells *in vitro*. The method comprises the steps of preparing a culture of neural precursor cells in a serum-free medium; culturing the neural precursor cells in the presence of a first mitogen, where the first mitogen is selected from the group consisting of aFGF, bFGF, EGF, TGF α and combinations thereof; contacting the cells with an agent capable of being taken up by the cells and capable of expressing a c-myc gene and a selectable marker; further culturing the cells in a medium containing the first mitogen and a second mitogen, where the second mitogen is selected from the group consisting of aFGF, bFGF, EGF, TGF α , serum and combinations thereof, with the proviso that the second mitogen is other than the first mitogen; and collecting c-myc treated cells and co-culturing them with feeder cells free of the selectable marker and capable of supporting survival of the c-myc treated cells in a medium containing the first mitogen and the second mitogen, with the proviso that the second mitogen is other than the first mitogen.

In a preferred embodiment of this method, the c-myc gene is fused with other DNA elements, where the other DNA elements comprise at least one element selected from the group consisting of a ligand binding domain for an estrogen receptor, an

androgen receptor, a progesterone receptor, a glucocorticoid receptor, a thyroid hormone receptor, a retinoid receptor, and an ecdysone receptor.

In another preferred embodiment of this method, the medium containing the first mitogen and the second mitogen further comprises a myc-activating chemical selected from the group consisting of β -estradiol, RU38486, dexamethasone, thyroid hormones, retinoids, and ecdysone.

In a more preferred embodiment of this method the mammalian neural precursor cells are derived from a human. In another more preferred embodiment of this method, the mammalian neural precursor cells are derived from an *in vitro* culture of pluripotent embryonic stem cells.

The present application also reveals a cell line produced by this method. In a preferred embodiment of this cell line, the cells maintain a multipotential capacity to differentiate into neurons, astrocytes and oligodendrocytes. In other preferred embodiments of this cell line, the cells maintain a bipotential capacity to differentiate into neurons and astrocytes or into astrocytes and oligodendrocytes.

In more preferred embodiments of this cell line the cells maintain a unipotential capacity to differentiate into neurons or into astrocytes.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1. Arrangement of pMycER retrovirus plasmid. A linearized EcoR1 fragment containing the human c-myc gene fused to the ligand binding domain of the human estrogen receptor gene (Eiler et al., 1989, Nature 340: 60-68) was ligated downstream of the 5'LTR of pLXSN retroviral expression plasmid (Clontech). The final construct also contains a selectable marker, the neomycin resistance gene, Neo^r , under the SV40 promoter, P_{SV40} .

FIG. 2. Growth capacity of MycER-modified human CNS stem cells. In order to measure the growth rate and capacity, a MycER-modified human CNS stem cell line pool (HK18.2) derived from 18-week old human fetal cortical tissue was continuously expanded in culture for approximately 80 days. At each passage (solid circle), the cells were harvested, counted, and a fraction replated into new plates. This process was repeated for 12 passages. By dividing the increased cell number from the initial seeding density to the time of harvest by the duration of the culture per passage, an approximate doubling time was estimated (open triangle). The dotted line across the graph represents the averaged doubling time for the entire culture period. Cumulative expansion of the cells was calculated by multiplying the multiples of increased cell number at each passage and expressed as "Cumulative Fold-Expansion" over the initial cell number at day 0. The initial starting cell number at day 0 was 5.0×10^6 cells.

FIG.3. Stability of neuronal differentiation of MycER-modified human CNS stem cells.

A. Unmodified CNS stem cells differentiated and immunostained with anti-MAP2ab antibody;

B. Unmodified CNS stem cells differentiated and immunostained with anti-TH antibody;

C. MycER modified human cortical cells at passage 4, differentiated and immunostained with anti-MAP2ab antibody viewed at low magnification;

D. MycER modified human cortical cells at passage 4, differentiated and immunostained with anti-MAP2ab antibody viewed at high magnification;

E. MycER modified human cortical cells at passage 4, differentiated and immunostained with anti-TH antibody viewed at low magnification;

F. MycER modified human cortical cells at passage 4, differentiated and immunostained with anti-TH antibody viewed at high magnification;

G. MycER modified human cortical cells at passage 4, differentiated and immunostained with anti-GABA antibody viewed at low magnification;

H. MycER modified human cortical cells at passage 4, differentiated and immunostained with anti-GABA antibody viewed at high magnification;

I. MycER modified human cortical cells at passage 9, differentiated and immunostained with anti-MAP2ab antibody viewed at low magnification;

J. MycER modified human cortical cells at passage 9, differentiated and immunostained with anti-MAP2ab antibody viewed at high magnification;

K. MycER modified human cortical cells at passage 9, differentiated and immunostained with anti-TH antibody viewed at low magnification; and

L. MycER modified human cortical cells at passage 9, differentiated and immunostained with anti-TH antibody viewed at high magnification.

FIG. 4. Stability of neuronal differentiation. MycER-modified human cortical cell lines were differentiated at passage 4 and at passage 11. The number of neurons immunostained for MAP2ab or TH proteins were quantified and their proportions over the total cells are reported.

FIG. 5. MycER modified neuronal progenitors.

A. MycER-modified rat striatal progenitors immunostained with anti-tau antibody;

B. Morphology and arrangement of tau+/TuJ1- neuronal progenitors, immunostained with anti-tau antibody;

C. Morphology and arrangement of tau+/TuJ1+ neuronal progenitors, immunostained with anti-tau antibody; and

D. Morphology and arrangement of tau+/TuJ1+ neuronal progenitors of **C**, immunostained with anti-TuJ1 antibody.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT

Neural cells in culture are highly plastic. Even a brief exposure to suboptimal culture conditions such as serum can have subtle yet significant long-term effects on

the phenotype of the cells. Yet, almost all of the reported neural cultures employ serum as the primary source of mitogen. We and others have demonstrated that, in order to preserve the intrinsic differentiation potential of stem cells and other cells, it is critical to reduce the exposure of the cells to serum and that well-defined growth factors, particularly bFGF and/or epidermal growth factor, EGF, in a serum-free medium can proliferate a variety of different cell types in a single culture (Johe, U.S. Patent 5,753,506; Weiss et. al., U.S. Patent 5,851,832).

In the absence of a particular molecular marker for each of the cell types, isolating potentially thousands of distinct neural cell types that may exist in a single culture had not been feasible. In the previous works, we have described the methods and compositions of distinct CNS stem cell populations that give rise to a variety of different neurons in culture. Here, a reproducible and efficient method utilizing over-expression of the c-myc gene to stabilize the differentiation potentials of neural cells and to isolate stable clonal cell lines is described.

With continuous passage, CNS stem cells gradually lose their capacity to differentiate into neurons, thus becoming glial progenitors. The conditions which accelerate this process include high cell density during proliferation, poor attachment of the cells on extracellular matrix coated surface, and exposure to glia-promoting factors such as CNTF (ciliary neurotrophic factor), LIF (leukemia inhibitory factor), BMPs (bone morphogenic factors) and serum. In order to overcome this instability of neuronal differentiation capacity of CNS stem cells, we have introduced into the cells a cellular proto-oncogene, c-myc, whose activity can be regulated by the presence or absence of an extracellular molecule, β -estradiol.

Human and rat CNS stem cells harboring the fusion gene were grown in the continuous presence of mitogens and β -estradiol in the culture medium. Growth of the cells were significantly more robust, exhibiting faster mitotic rate, resistance to spontaneous differentiation, and much greater overall stability during the expansion. The cells showed no sign of neoplastic transformation or anomalous growth pattern or morphology. Upon withdrawal of the mitogens and β -estradiol, the cells initiated differentiation promptly and gave rise to heterogeneous morphologies characteristic of neurons and glia. Neuronal differentiation was efficient, exhibiting molecular expression patterns, localization of neurons-specific proteins, and cell morphologies and behaviors essentially indistinguishable from the parental unmodified CNS stem cells.

The neuronal population consisted of various neurotransmitter phenotypes, including the tyrosine hydroxylase-positive dopaminergic phenotype in 10-20% of the neurons. Such neuronal differentiation capacity was stable through over 60 cell doublings resulting in at least 1×10^{18} -fold increase in the number of neurons and glia derived from the stem cells. Thus, the genetic modification and the stem cell culture method described here enable the stable isolation of practically unlimited numbers of CNS stem cells from all regions of the developing mammalian brain, each CNS stem cell clone giving rise to potentially distinct neuronal subtypes in unlimited numbers. The result, then, is a library of mammalian neurons, including human, with distinct

molecular/genetic repertoires representing the diverse cellular phenotypes of the mature brains.

Examples

Construction of c-Myc-estrogen receptor expressing retrovirus

A retroviral vector containing the neomycin-resistance gene under the SV40 promoter was linearized with EcoRI and ligated with the EcoRI-fragment of the DNA encoding a fusion gene of human c-myc cDNA and human estrogen receptor cDNA (Eiler et al., 1989, Nature 340: 60-68). The fusion gene was placed under the regulation of MMLV long-terminal repeat sequence (LTR). The overall arrangement of the final retroviral vector, pMycER, is shown in Figure 1.

Generation of a producer cell line

To establish a cell line stably producing the MycER retrovirus, an amphotropic packaging cell line was transfected with pMycER plasmid. Stable clones were selected with G418 (1 mg/ml, Life Technology Inc., MD) for 4 weeks. Twenty clones were screened for high titer production against Hela cells according to standard procedure. A cell line, MycER.10, with a retroviral titer of 10^5 pfu/ml as measured by infection of rat striatal stem cells was selected for subsequent experiments.

Infection of rat and human CNS stem cells

Rat and human CNS stem cells were prepared according to previously reported procedure (U.S. Patent 5,753,506). Passage 1 cells were plated at 0.5×10^6 cells per 100 mm plate and grown for three additional days in serum-free N2 medium plus 10ng/ml bFGF. MycER.10 cells were grown in DMEM/10% fetal bovine serum to 50-75% confluence, subsequently rinsed three times with DMEM, and incubated for 4-16 hours in a retrovirus collection medium (IFM). IFM consisted of the standard N2 components (25mg/L human recombinant insulin, 100mg/L human apotransferrin, progesterone, putrescine, sodium selenite) in DMEM plus 10ng/ml bFGF and 1 μ g/ml human plasma fibronectin (hFN). IFM containing the retrovirus was clarified by two centrifugations at 1400 rpm and 3000 rpm. The supernatant was mixed with fresh N2 at a 1:1 ratio with fresh bFGF and hFN at 10ng/ml and 1 μ g/ml final concentrations, respectively, and applied to the 50-75% confluent CNS stem cell culture. The infection period was typically 6 hours. Human CNS stem cells were infected for 1-3 times over a 2-3 day period to compensate for their slow mitotic rate. Subsequently, the cells were rinsed three times with Ca²⁺-, Mg²⁺-free Hank's balanced saline solution (HBSS), passaged, and further expanded in N2 plus 10ng/ml bFGF.

Selection of MycER-expressing CNS stem cells

The CNS stem cells with stable incorporation of MycER retrovirus were passaged 1-2 days after the infection, replated at 0.5×10^6 cells per 100 mm plate, and

selected from 2 days after the infection with 0.1-0.2 mg/ml G418, pH 7.4. The complete, optimal growth medium (IGM) was composed of DMEM/F12 (1:1), 25mg/L human recombinant insulin, 100mg/L human apotransferrin, progesterone, putrescene, sodium selenite, 10ng/ml bFGF, 0.2 μ M β -estradiol, 0.1mg/ml G418, and 10ng/ml EGF or 1% fetal bovine serum. Fresh bFGF (10ng/ml, final concentration) was added daily and medium was changed once every two days. The cells were passaged at approximately 50-75% confluence by rinsing three times with HBSS and trypsin (1x) treatment. Trypsin activity was stopped by adding soybean trypsin inhibitor (1mg/ml final concentration).

Isolation of clones

At the end of the G418 treatment for 14 days, the cells were passaged and replated approximately 200-1000 cells per 100 mm plate. Within 24 hours post plating, well-isolated single cells were marked with 3 mm circles on the bottom of the culture plate. Sometimes, the complete culture medium was mixed with an equal volume of the medium conditioned by the same cells at high cell density to enhance cell survival. Marked clones were picked with the aid of cloning rings and by trypsin treatment. Individual clones were expanded as the mass culture and stored frozen.

Almost all of the clones generated this way eventually assumed glial morphology and failed to differentiate into neurons, even though the culture conditions are identical as those for the high density culture. Thus, it became apparent that the MycER-modified cells in the presence of serum required a relatively high cell density in order to maintain their native differentiation potentials and survive. Thus, the cell density was maintained in the range of 0.5×10^6 to 1.0×10^6 cells per 100 mm plate by supplementing the clonal density of MycER-expressing cells with unmodified primary stem cells. By maintaining antibiotic selection over 5-8 days with 0.1 mg/ml G418, the feeder cell population was gradually killed while permitting local cell density of G418-resistant MycER cells to gradually rise so as to sustain their optimal growth. The antibiotic selection was maintained throughout subsequent expansion to ensure all remaining cells were MycER-modified cells.

In addition to neural stem cells, immature glial cells and mature astrocytes of both human and rat origin were effective. Fibroblasts were also useful, but more difficult to manage because their rapid proliferation rate and their high tolerance to G418. Neurons may also be useful, but their post-mitotic nature rendered them much more resistant to G418. Non-mitotic fibroblasts and other non-neural cells which had been gamma-irradiated or treated with mitotic inhibitors such as arabinoside C or mitomycin may also be effective in supporting the c-myc-modified neural cells.

Differentiation and characterization of the mitotically enhanced CNS stem cells

CNS stem cells stably expressing MycER were differentiated by plating the cells at 100,000 cell/cm² or higher cell density and replacing the growth medium with N2

without bFGF, without serum, and without β -estradiol. Typically, the cells were allowed to differentiate for 6-30 days before immunohistochemical analysis.

Results

1. Search for additional factors enhancing mitotic capacity of CNS stem cells

The doubling time for human CNS stem cells in N2 medium with bFGF as the sole mitogen is approximately 60 hours which is markedly slower than 24 hours for rat CNS stem cells under the identical culture conditions. This may be due to a species difference in certain cell-autonomous properties of the cells such as a difference in DNA replication rate or in other mitotic phases, G1 or G2, of the cell cycle. We investigated other factors to accelerate the mitotic rate of human CNS stem cells.

Many purified recombinant human growth factors were tested for the ability to enhance the mitogenic activity of bFGF. The mitotic rate of human CNS stem cells was assessed by measuring the proportion of the cells which have incorporated the mitotic label, bromodeoxyuridine (BrdU), during a 24-hour period. Each growth factor was supplied daily to the culture in addition to bFGF. Only the combination of bFGF + EGF and bFGF + TGF α (transforming growth factor-alpha) accelerated beyond the bFGF-induced mitotic rate of the human CNS stem cells. In both conditions, the doubling time of the cells increased 1.5 times to 40 hours over bFGF alone. Combination of bFGF and fetal bovine serum at 1% or at 10% also accelerated the BrdU-incorporation rate of bFGF-induced mitotic rate of CNS stem cells to a similar rate.

2. Resistance of CNS stem cells against spontaneous differentiation

Although EGF, TGF α , 1%FBS, or 10%FBS plus bFGF increased the mitotic rate of human CNS stem cells, even under these conditions, CNS stem cells were susceptible to spontaneous differentiation at near-confluent cell density and also prone to drift toward glial progenitor states with multiple passages. In order to provide enhanced mitotic capacity and greater stability to the neuronal differentiation capacity of CNS stem cells, we constructed a retroviral vector expressing a fusion protein of human c-myc and human estrogen receptor genes under the MMLV long terminal repeat (Figure 1).

Dividing mammalian CNS stem cells were infected by the amphotropic retrovirus with high efficiency and the resulting cells selected by their resistance to G418 treatment. The transcriptional activity of c-myc in the fusion protein (MycER) was regulated by the presence or absence of the estrogen receptor ligand, β -estradiol, in the culture medium. Moreover, the promoter activity of the long terminal repeat is shut down during CNS stem cell differentiation into neurons, effectively eliminating MycER transcript. Combination of the withdrawal of mitogens, absence of β -estradiol, and limited transcription activity of LTR resulted in an efficient constitutive differentiation of CNS stem cells in a manner indistinguishable from the unmodified parental cells.

CNS stem cells from various regions and developmental stages of human fetal brains were infected at passage 1 with the MycER-expressing retrovirus. Infected cells were selected by G418 resistance and expanded in N2B medium (N2 without phenol red) containing bFGF, 10ng/ml EGF or 1% FBS, and β -estradiol. Expression of MycER itself did not cause a significant change in the mitotic rate of the cells; however, the addition of EGF and/or FBS significantly increased the mitotic rate and enhanced the overall stability of the culture. The cells proliferated robustly, maintained stable morphologies over many successive passages, and sustained their multipotentiality without spontaneous differentiation even at nearly confluent cell density. Upon replacement of the growth medium with N2B without any mitogen and without β -estradiol, the stem cells promptly differentiated to give rise to neurons, astrocytes and oligodendrocytes.

3. Expansion capacity of the mitotically enhanced CNS stem cell lines

To ascertain the extent of the mitotic and differentiative capacity of the MycER-modified human CNS stem cell line, the cells were expanded continuously for 80 days in culture and through 12 passages since the infection event. During this period, the cell yield at each passage was measured to quantify the actual arithmetic increase in cell number and to determine the stability of the mitotic rate over time (Figure 2). Overall, the cells went through approximately 54 doublings which resulted in 10^{15} -fold increase in the cell number. The doubling time of the cells was remarkably constant at about 40 hours per mitosis, which is unchanged from that of the parental primary human CNS stem cells (Figure 2).

The same human CNS stem cell preparation was also subjected to MycER-retrovirus infection and grown in bFGF alone or in bFGF and EGF. In bFGF alone as the mitogen, the MycER-expressing CNS stem cells exhibited enhanced mitotic capacity over the unmodified cells, but yet showed far less proliferative capacity than in bFGF + 1% FBS. As with the unmodified parent cells, the MycER cells also retained a 60 hour doubling time in bFGF alone. On the other hand, in bFGF + EGF, the MycER expressing stem cells displayed increased mitotic rate, increased mitotic capacity, increased stability of neuronal differentiation capacity, and were resistant to spontaneous differentiation quite similar to the bFGF + 1% FBS condition. In the absence of MycER expression, the same three conditions yielded a similar pattern of growth, but with less stability. Significantly, the bFGF + 1%FBS condition, although resulting in more efficient cell growth, inevitably led to the loss of the neuronal differentiation capacity. This demonstrates that the constitutive c-myc function in these cells is subtle: It provides more stable multipotentiality and enhanced mitotic capacity but not an overt mitogen-independence or transformation.

These effects of the constitutively active c-myc could also be extended to CNS stem cells from all regions of rat and human fetal brains.

4. Neuronal differentiation of MycER-enhanced CNS stem cells

The MycER-enhanced CNS stem cells were differentiated by withdrawing the mitogens and β -estradiol from the medium and without addition of exogenous factors. Divergence of neuronal and glial morphologies began to occur within two days. By the third day, neuronal morphologies were clearly distinguishable. The neurons continued to mature into fully functional neurons over the next 3-5 weeks.

The differentiated cultures from the MycER-enhanced human CNS stem cells at different passages were analyzed by immunohistochemistry with a variety of different cell-type specific antibodies. At 10 days of differentiation, approximately 50% of the total cells expressed MAP2c, tau, and tubulin IIIb proteins, all relatively early markers of neuronal differentiation. Approximately 20-30% of the total cells expressed the mature markers of neurons, MAP2a and MAP2b proteins. Various neurofilament antibodies revealed a similar proportion of neurons. Of the neurons, approximately 70% were GABA-positive. A similar proportion of neurons was also calretinin-positive. Approximately 10-20% of the neurons expressed tyrosine hydroxylase (TH), the key biosynthetic enzyme for dopamine. All of the immunopositive neurons were of typical neuronal morphologies and did not co-express the glial marker, GFAP. Thus, MycER-enhanced cell lines differentiate to generate a high proportion of neurons exhibiting various neurotransmitter phenotypes.

The proportions and neurotransmitter phenotypes of the neurons were stable through many successive passages (Figure 3A-L). Throughout 54 stem cell doublings, there was no degradation of the neuronal differentiation capacity in both the proportion of neurons as well as in the various neurotransmitter phenotypes generated.

5. Region-specific stem cell lines

The serum-free culture condition used for isolation of neural stem cells permitted stable inheritance of regional identities and their related neurotransmitter phenotypes through multiple cell divisions. This implies that the stem cells in the culture, although they are uniform in their ability to differentiate into neurons and glia, may be extremely diverse. Thus, if each stem cell in the beginning of the culture could be immortalized in its native state and if this method was efficient to sample thousands of stem cells in a single dish, then the diverse neuronal phenotypes might be permanently "captured" in the form of cell lines.

The genetic modification of neural stem cells with c-myc resulted in robust, highly reproducible and in a stable cell culture system. The modification process itself is quite efficient, yielding 5,000 to 50,000 independent clones per retrovirus infection over a two day process. This could be easily scaled up by increasing the retrovirus particles or increasing the number of target cell density, if needed.

In order to ascertain whether over-expression of c-myc has an impact on differentiation capacity of neurotransmitter phenotypes, stem cells from many different regions of the fetal brains of rat and human were modified by the MycER retrovirus. These regions included cortex, septum, hippocampus, midbrain, hindbrain, striatum,

and spinal cord. Multiple examples of cortical, midbrain, and spinal cord cultures from several different gestational ages were examined to assess the reproducibility of the method. In all cases, the resulting pools of independent clones generated highly reproducible ratios of neurons to glia. As expected, morphologies, antigenic profiles of the neurons, and their relative ratios were also distinct from cell lines of one region versus another.

Thus, when several pools of cell lines from midbrain tissues of 8 week human fetuses were examined, approximately 0.1% of total cells were consistently TH-positive dopaminergic neurons, which is also the proportion found in the unmodified stem cell cultures. Clonal analysis revealed that the TH expression was clonally restricted. That is, a majority of the clones did not contain neurons expressing TH. Of those that did, the proportion was variable from one clone to the next. Several pools of cell lines from cortical tissues of 17-20 week old human fetuses were also examined. Interestingly, all of the cortical lines gave rise to significantly increased TH-positive neurons compared to the unmodified stem cells. The proportion of TH-positive neurons was 2-4% of the total cells. Subsequently clonal analysis revealed a similar pattern in the distribution of the TH-positive neurons. The majority had none, while those that generate TH were present in variable proportions. This pattern had also been observed with clones of unmodified stem cells from several different regions and with several different antigenic markers.

Cell lines from spinal cord of 6-10 week old human fetuses were also established. The pattern of neuronal differentiation was the same as from other regions, although their stem cell morphology and growth characteristics were distinct.

Thus, the genetic modification of neural stem cells with c-myc does not alter their intrinsic differentiation capacities. In all of the cell lines through extensive continuous culture periods, no evidence of tumor formation or other abnormal transformation was noted. Upon karyotype analysis of one pool of human cortical cell lines at passage 14, a normal diploid chromosome pattern with no aberrant rearrangement was observed. Thus, regulation of mitotic capacity by c-myc, which is a cellular gene normally present in every eukaryotic cell and a well-known key regulator of cell cycle machinery, is not oncogenic and provides significant advantages over other methods using viral oncogenes such as v-myc or SV40 large T antigen.

6. Other cell types

The genetic modification with c-myc can be made at any time during the culture period. Since the expression of c-myc itself is not mitogenic, i.e., non-transforming, a culture condition which promotes the proliferation of a particular neural precursor population is a prerequisite. Purified growth factors such as aFGF (acidic fibroblast growth factor), bFGF, EFG and TGF α can proliferate a variety of different neural cell types. Although most of the descriptions above were on multipotent CNS stem cells as one predominant population, several different cell types were observed during clonal analysis.

One significant population was a population of bipotential progenitor clones which, upon differentiation, gave rise to neurons and astrocytes with apparent absence of oligodendrocytes. These bipotential progenitors were quite similar to the multipotential stem cells in their morphology during growth. The differentiation pattern was also similar giving rise to about 50% neurons and 50% astrocytes. Thus, key defining difference between the two populations is the absence of oligodendrocytes in differentiated cultures.

A second cell population arising from c-myc modification of primary neural cultures was a population of unipotential neuronal progenitor clones which consisted only of neurons. These neuronal progenitor clones were of smaller clone size and assumed distinct, immature neuronal morphologies during proliferation and expressed tau proteins and/or beta-tubulin III. Examples are provided in Figure 5. Two distinct cell types were observed (FIG 5A). One type was small-bodied cells with very short single process stemming out from the cell body, which grew in tight clusters. These cells were immunoreactive with anti-tau antibody but not with anti-tubulin IIIb antibody while dividing (FIG 5B). The other type of cells were cells with distinctively elongated neurites without extensive branching, which grew in a smaller, more scattered pattern suggesting higher migratory capacity. In contrast to the first type, they were also immunoreactive with both anti-tau antibody and with anti-tubulin IIIb antibody (FIG 5C and 5D, respectively). Often times, the second cell types were found near or intermixed with the first cell type suggesting that they are two stages of a single continuous lineage—committed neuronal progenitors, with tau+/TuJ1+ state being the more mature state.

A third cell population arising from c-myc-modified neural cell culture was a population of clones consisting of glia only. Most of these clones were astrocytic with little or no oligodendrocytes.

These results indicate that many neural precursor lineages respond similarly to the over-expression of c-myc. In addition to primary neural cultures prepared from nervous system tissues of mammals, recent advances in embryonic stem cell cultures indicate that various neural precursors form *in vitro* during differentiation of totipotent or pluripotent embryonic stem cells and cell lines maintained in culture for long term (Renoncourt et al., Mech. Dev. (1998) 78, 185; Svendsen et. al., Trends Neurosci. (1999) 22, 357; Brustle et. al., Science (1999) 285, 754.). These cultures can generate nesting-positive neural precursor cells which can then be transferred to serum-free medium and subsequently expanded with bFGF and/or EGF for short term. Long-term, mass expansion has not been feasible since the initial neural precursor formation is inefficient. However, by utilizing the genetic modification method with c-myc gene described here, those transient neural precursors may be turned into stable cell lines.

Neural precursors including multipotential neural stem cells can be isolated from adult brains and can be cultured in serum-free conditions. However, this process is inefficient, resulting in only a small number of proliferative cells. However, with the

transfer of c-myc gene as described here, one can establish stable cell lines from such small number of cells obtained from neural tissue biopsies.

c-myc is involved in many different cellular processes such as apoptosis in addition to cell cycle regulation. c-myc has been used previously to transform cells of non-neural origins. However, these previous studies were done with already stable cell lines such as 3T3 fibroblast cell lines and to produce neoplastic state of the cell lines, which had been selected based on spontaneous chromosomal aberrations which conferred mitogen-independence. Other studies tried to use an immortalization process to turn post-mitotic neurons to re-enter the cell cycle.

CNS stem cells are already mitotic and the mitogenic culture conditions for a long-term expansion of up to 30 cell doublings has already been established. Thus, our objective has been to increase the expansion capacity well beyond the 30 cell doublings at least up to the beginning of senescence which is thought to occur between 60 and 80 cell cycles. Sixty cell doublings represent an 1×10^{18} -fold increase in cell number which is large enough for screening one million chemical libraries, each consisting of 500,000 compounds or large enough to provide cell therapies for 50 billion Parkinson's patients. The key concept has been to find a "gentle" modification of the cells so as not to disrupt their intrinsic neuronal differentiation capacity while providing an enhanced growth capacity under the culture conditions established for primary CNS stem cells.

Increasing the concentration of active c-myc protein leads to the generation of stable human CNS stem cell lines. This effect occurs not by overtly deregulating mitotic and differentiation parameters of the cell cycle but by providing resistance to autocrine and paracrine factors that induce restriction of multipotentiality toward a glial progenitor state. The consequence is not an oncogenic transformation of the cells but rather a stabilization of the cell growth. Thus, endogenous signals which trigger the differentiation, such as those present at confluent cell density, are still effective. The cell division is still dependent upon the supply of proper exogenous mitogens such as bFGF and/or EGF and/or serum. Differentiation of the stem cell lines to mature functional neurons is as efficient at the end of the 60-cell doublings as in the unmodified primary cells. A variety of neurotransmitter phenotypes and their relative proportions are maintained throughout the expansion.

The c-myc activity in these examples was controlled by constructing a chimeric protein of c-myc fused to a fragment of estrogen receptor protein (Eiler et. al., Nature (1988) 340, 60). The intended role of the estrogen is to provide a control over the amount of functionally active c-myc induced in the cell. The estrogen receptor portion of the chimeric protein is activated when it binds with a cell-permeable agonist or antagonist such as β -estradiol or tamoxifen.

Most members of the nuclear receptor superfamily act similarly in that cell-permeable ligands diffuse through the plasma bilayer and bind to their receptor which is then transported to the nucleus as a complex and induces a variety of transcription

related events. The ligand binding domain of these nuclear receptor proteins and their ligands can substitute for the estrogen receptor and β -estradiol in order to regulate functions of the fused c-myc protein moiety. Examples of such nuclear receptors are glucocorticoid receptor, progesterone receptor, androgen receptors, vitamin D receptor, thyroid hormone receptors, retinoic acid receptors, and ecdysone receptor. Each of these receptors can be activated intracellularly by adding to the culture medium its appropriate ligands. Examples of the ligands are steroid hormones such as glucocorticoid or dexamethasone, thyroid hormones, retinoids such as retinoic acids, vitamin D, and the insect molting hormone, ecdysone, as well as their synthetic analogs designed to act on the respective receptors. All of these compounds are small, hydrophobic molecules which can traverse the cell membrane once supplied extracellularly.

Some receptor-ligand systems are better suited than others for the purpose of regulating the over-expressed c-myc. For instance, for the purpose of transplanting c-myc-modified cells into tissues as a treatment of a disease, it would be desirable that the c-myc-receptor chimeric protein should not respond to endogenous physiological ligands. The c-myc-estrogen receptor described here has the disadvantage that potentially high level of estrogen present in female patients may have unexpected effects on the cells. In another instance, the ligands used to control the c-myc activity in culture may have their own unrelated effects on endogenous receptors. Thus, an ideal receptor-ligand system is one in which the receptor moiety of the fusion protein does not recognize the endogenous ligand and in which the ligand is a synthetic compound which has no adverse effect on the cells. One such potential system is human progesterone receptor and its antagonist ligand, RU38486. It has been established that the ligand binding fragment of the human progesterone receptor does not respond to the endogenous ligand, progesterone but is sensitively activated by a synthetic analog of progesterone, RU38486, while RU38486 does not activate the endogenous full-length progesterone receptor (Wang et al., Proc. Natl. Acad. Sci. USA (1994) 91, 8180).

Thus, one enhanced c-myc expression system to produce stable cell lines would be to construct a plasmid in which the human c-myc gene is fused to the ligand binding domain of the human progesterone receptor with the C-terminal deletion of 12 amino acids, to cut out the fused DNA (c-mycPR), ligate to the retroviral plasmid, pLXSN, at downstream of 5' LTR, and to generate the intact retrovirus expressing the chimeric protein, c-myc-progesterone receptor (MycPR).

The commercial utilities of the mitotically enhanced CNS stem cells are: cell transplantation of the TH-positive dopaminergic neurons to treat Parkinson's disease; substrate for screening potential pharmacological compounds; a reproducible source of gene and protein levels of the cells influenced by a specific agent or protocol designed to represent/mimic a disease process; a reproducible source of novel genes and proteins; a reproducible source of neurons and glia for engineering of three dimensional tissues and neural prosthesis; a delivery vehicle of potentially therapeutic large molecule compounds such as NGF to treat Alzheimer's disease; and the starting

population to further derive *in vitro* various committed neuronal progenitor populations such as proliferative TH-expressing neuronal cells.

While the invention has been described in connection with what is presently considered to be the most practical and preferred embodiments, it is to be understood that the invention is not limited to the disclosed embodiments, but on the contrary is intended to cover various modifications and equivalent arrangements included within the spirit and scope of the appended claims. Thus, it is to be understood that variations in the present invention can be made without departing from the novel aspects of this invention as defined in the claims. All patents and articles cited herein are hereby incorporated by reference in their entirety and relied upon.

Claims

WHAT IS CLAIMED IS:

1. A method for producing stable cell lines of mammalian neural precursor cells *in vitro*, comprising the steps of:

- a) preparing a culture of neural precursor cells in a serum-free medium;
- b) culturing the neural precursor cells in the presence of a first mitogen, wherein said first mitogen is selected from the group consisting of aFGF, bFGF, EGF, TGF α and combinations thereof;
- c) contacting the cells with an agent capable of being taken up by the cells and capable of expressing a c-myc gene; and
- d) further culturing the cells in a medium containing the first mitogen and a second mitogen, wherein said second mitogen is selected from the group consisting of aFGF, bFGF, EGF, TGF α , serum and combinations thereof, with the proviso that the second mitogen is other than the first mitogen.

2. The method of claim 1, wherein the c-myc gene is fused with other DNA elements, wherein said other DNA elements comprise at least one element selected from the group consisting of a ligand binding domain for an estrogen receptor, an androgen receptor, a progesterone receptor, a glucocorticoid receptor, a thyroid hormone receptor, a retinoid receptor, and an ecdysone receptor.

3. The method of claim 1, wherein said medium containing the first mitogen and the second mitogen further comprises a myc-activating chemical selected from the group consisting of β -estradiol, RU38486, dexamethasone, thyroid hormones, retinoids, and ecdysone.

4. The method of claim 1, wherein the mammalian neural precursor cells are derived from a human.

5. The method of claim 1, wherein the mammalian neural precursor cells are derived from an *in vitro* culture of pluripotent embryonic stem cells.

6. A cell line produced according to the method of claim 1.

7. The cell line of claim 6, wherein the cells maintain a multipotential capacity to differentiate into neurons, astrocytes and oligodendrocytes.

8. The cell line of claim 6, wherein the cells maintain a bipotential capacity to differentiate into neurons and astrocytes.

9. The cell line of claim 6, wherein the cells maintain a bipotential capacity to differentiate into astrocytes and oligodendrocytes.

10. The cell line of claim 6, wherein the cells maintain a unipotential capacity to differentiate into neurons.

11. The cell line of claim 6, wherein the cells maintain a unipotential capacity to differentiate into astrocytes.

12. A method for producing stable clonal cell lines of mammalian neural precursor cells *in vitro*, comprising the steps of:

- a) preparing a culture of neural precursor cells in a serum-free medium;
- b) culturing the neural precursor cells in the presence of a first mitogen, wherein said first mitogen is selected from the group consisting of aFGF, bFGF, EGF, TGF α and combinations thereof;
- c) contacting the cells with an agent capable of being taken up by the cells and capable of expressing a c-myc gene and a selectable marker;
- d) further culturing the cells in a medium containing the first mitogen and a second mitogen, wherein said second mitogen is selected from the group consisting of aFGF, bFGF, EGF, TGF α , serum and combinations thereof, with the proviso that the second mitogen is other than the first mitogen; and
- e) collecting c-myc treated cells and co-culturing them with feeder cells free of the selectable marker and capable of supporting survival of the c-myc treated cells in a medium containing the first mitogen and the second mitogen, with the proviso that the second mitogen is other than the first mitogen.

13. The method of claim 12, wherein the c-myc gene is fused with other DNA elements, wherein said other DNA elements comprise at least one element selected from the group consisting of a ligand binding domain for an estrogen receptor, an androgen receptor, a progesterone receptor, a glucocorticoid receptor, a thyroid hormone receptor, a retinoid receptor, and an ecdysone receptor.

14. The method of claim 12, wherein said medium containing the first mitogen and the second mitogen further comprises a myc-activating chemical selected from the group consisting of β -estradiol, RU38486, dexamethasone, thyroid hormones, retinoids, and ecdysone.

15. The method of claim 12, wherein the mammalian neural precursor cells are derived from a human.

16. The method of claim 12, wherein the mammalian neural precursor cells are derived from an *in vitro* culture of pluripotent embryonic stem cells.

17. A cell line produced according to the method of claim 12.

18. The cell line of claim 17, wherein the cells maintain a multipotential capacity to differentiate into neurons, astrocytes and oligodendrocytes.

19. The cell line of claim 17, wherein the cells maintain a bipotential capacity to differentiate into neurons and astrocytes.

20. The cell line of claim 17, wherein the cells maintain a bipotential capacity to differentiate into astrocytes and oligodendrocytes.

21. The cell line of claim 17, wherein the cells maintain a unipotential capacity to differentiate into neurons.

22. The cell line of claim 17, wherein the cells maintain a unipotential capacity to differentiate into astrocytes.

Abstract

A systematic and efficient method for establishing stable neural stem cell lines and neuronal progenitor lines is described. The resulting cell lines provide robust, simple, and reproducible cultures of human and other mammalian neurons in commercially useful mass quantities while maintaining normal karyotypes and normal neuronal phenotypes.

FIG. 1

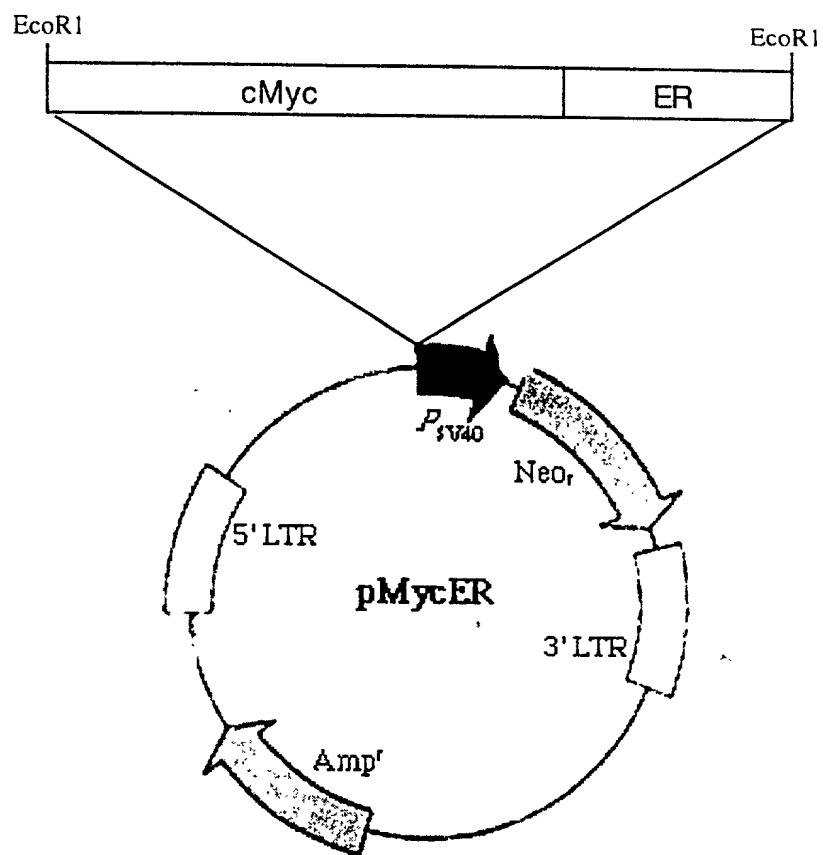


FIG. 2

Growth Capacity of MycER-modified Human CNS Stem Cells

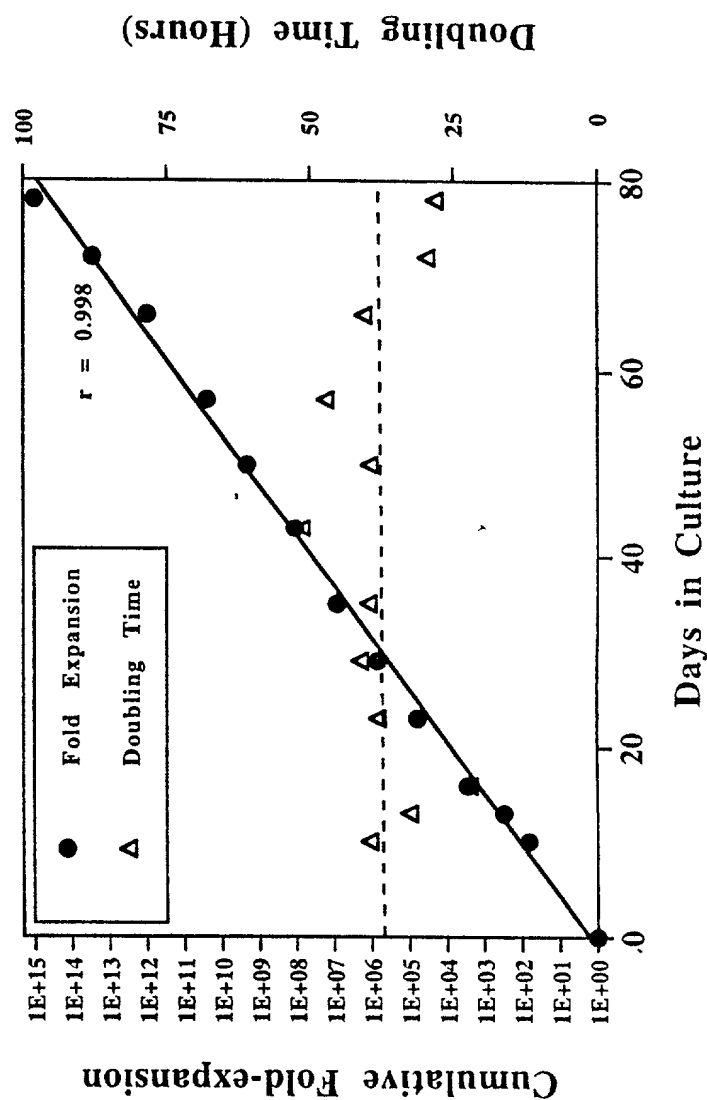


FIG. 3A

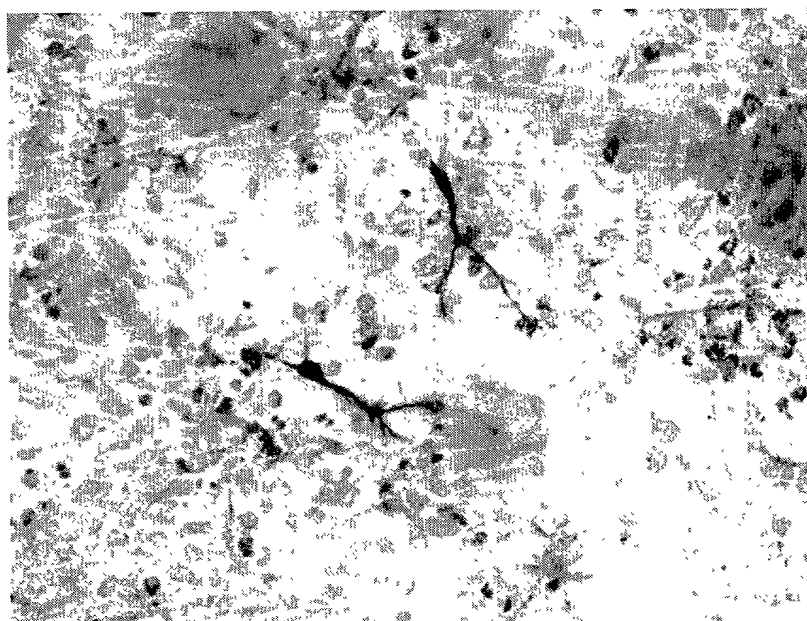
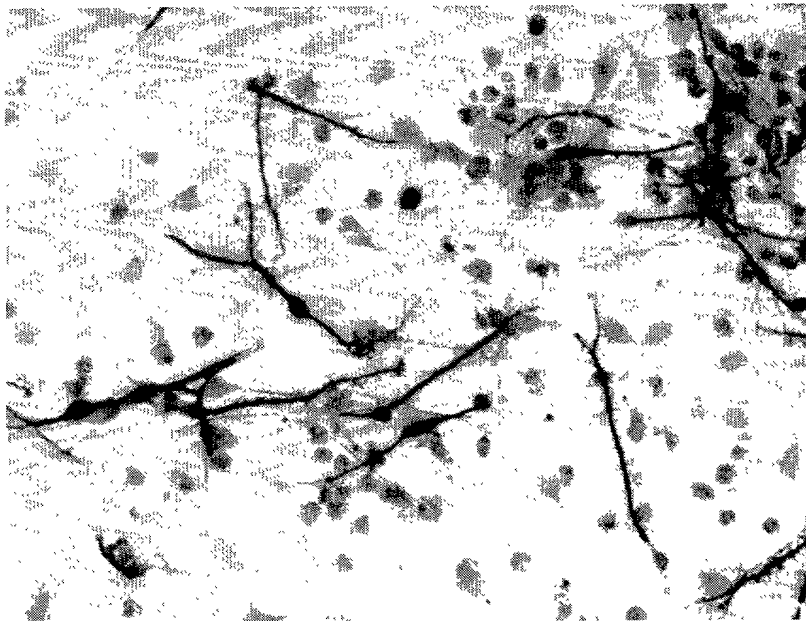


FIG. 3B

FIG. 3C

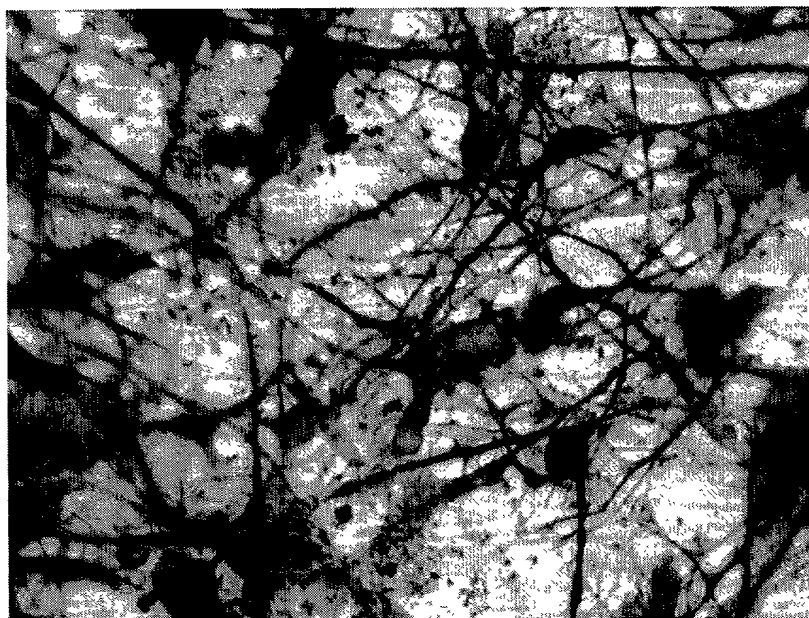
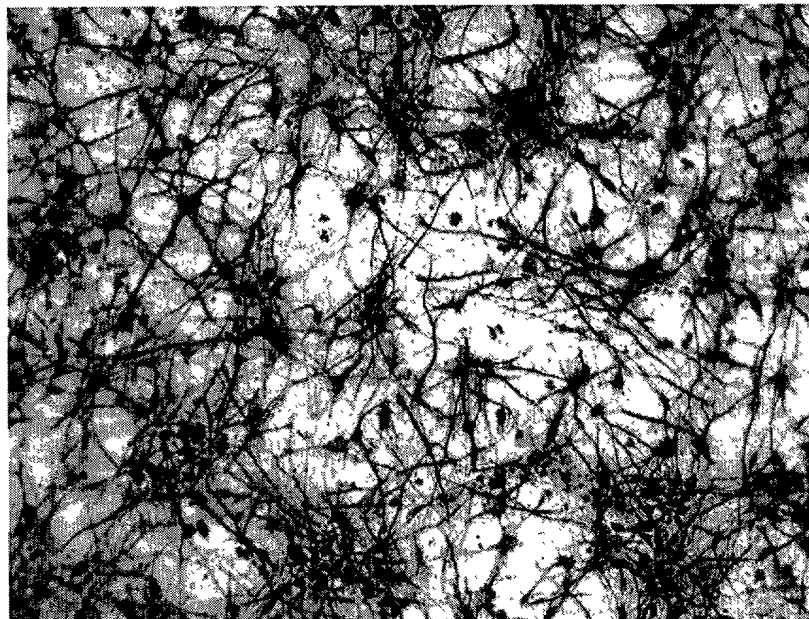


FIG. 3D

FIG. 3E

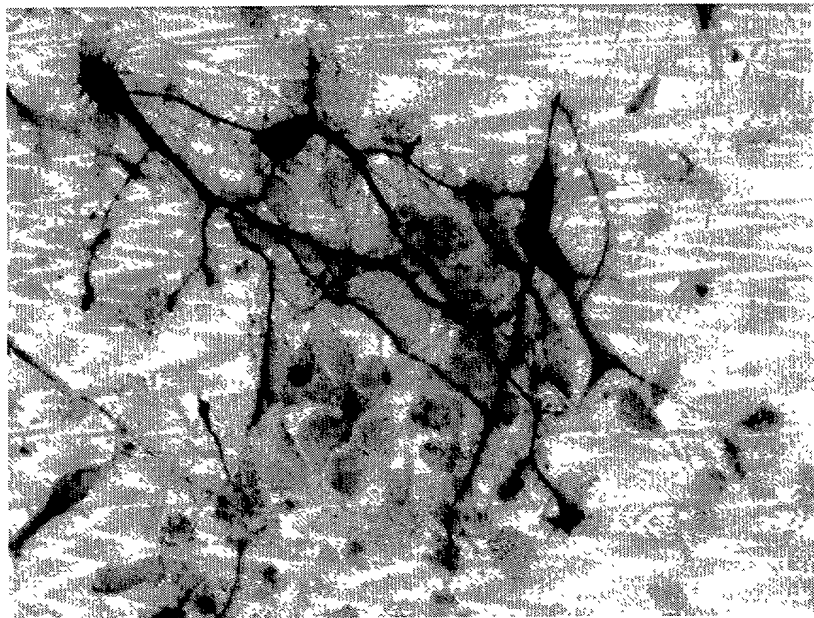
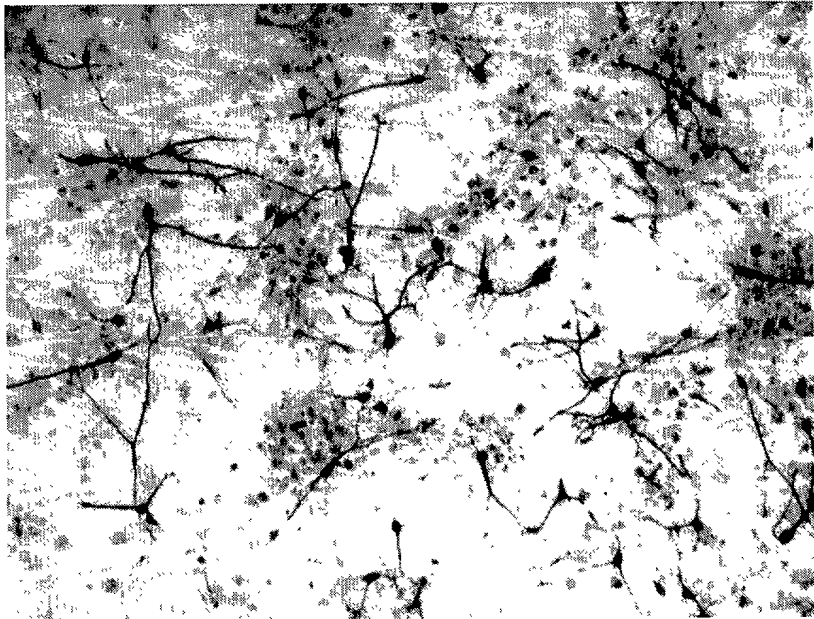


FIG. 3F

660250 26886E60

FIG. 3G

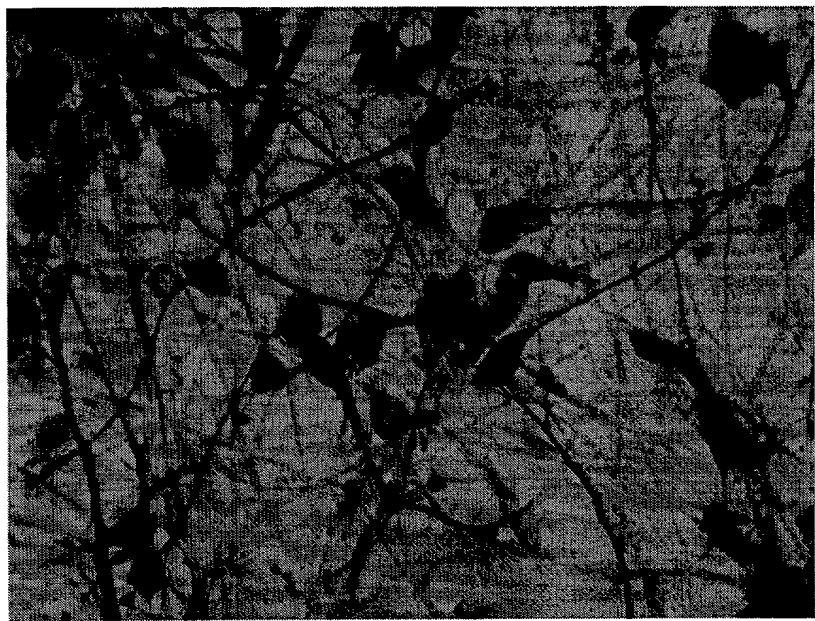
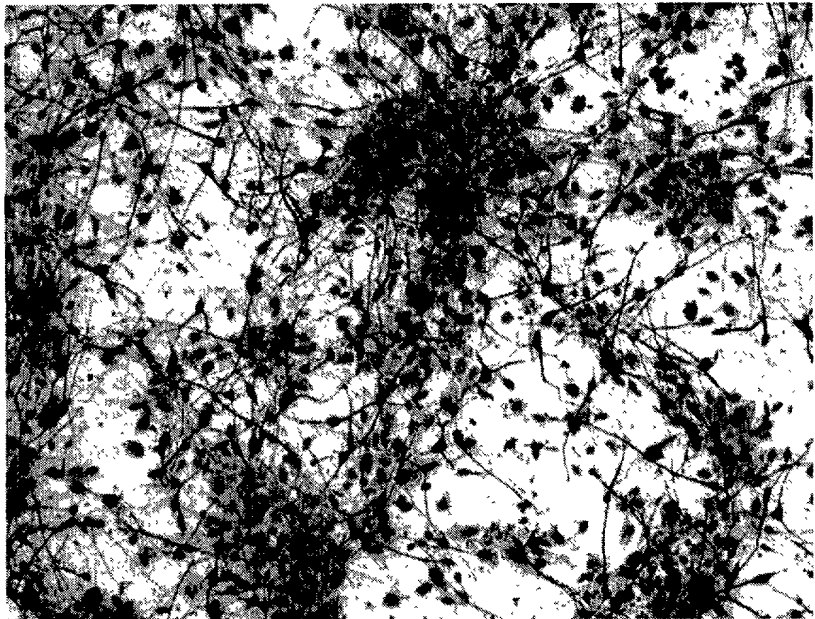


FIG. 3H

09398897 092099
560260 26886360

FIG. 3I

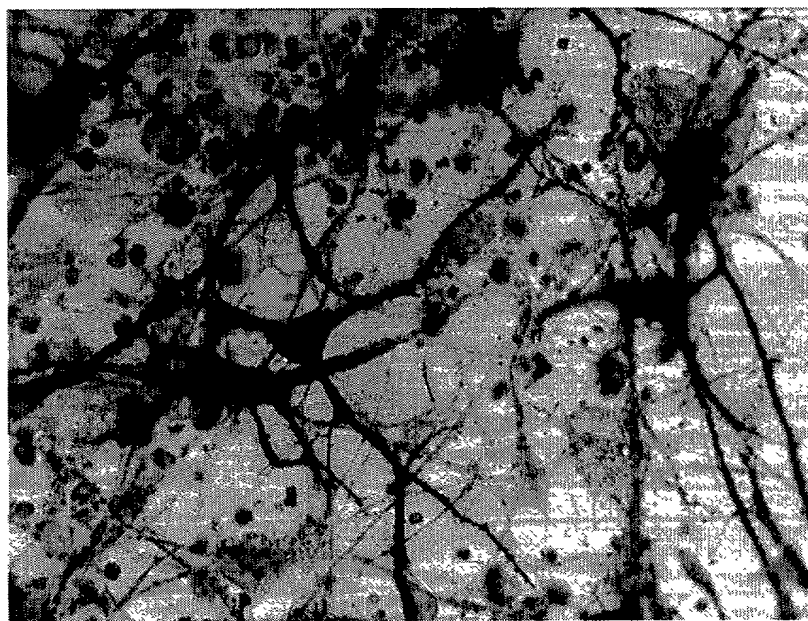
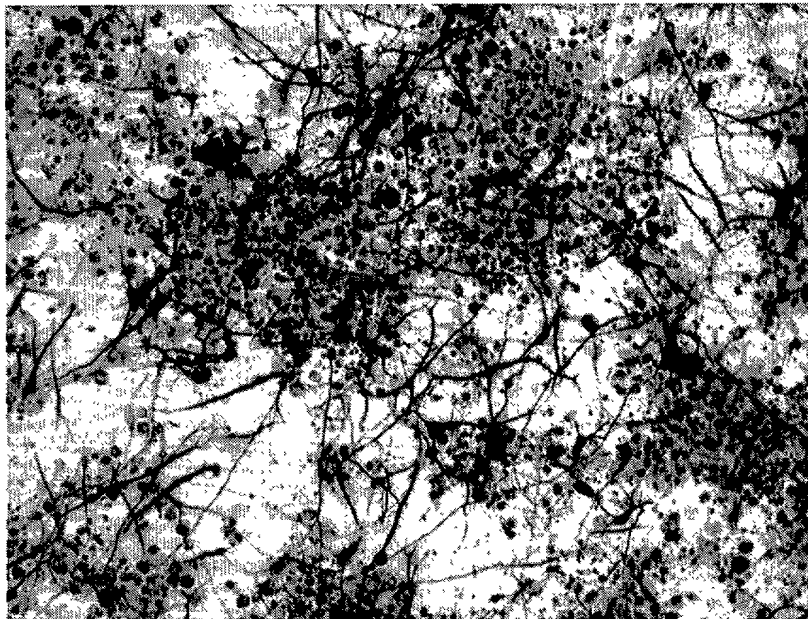


FIG. 3J

FIG. 3K

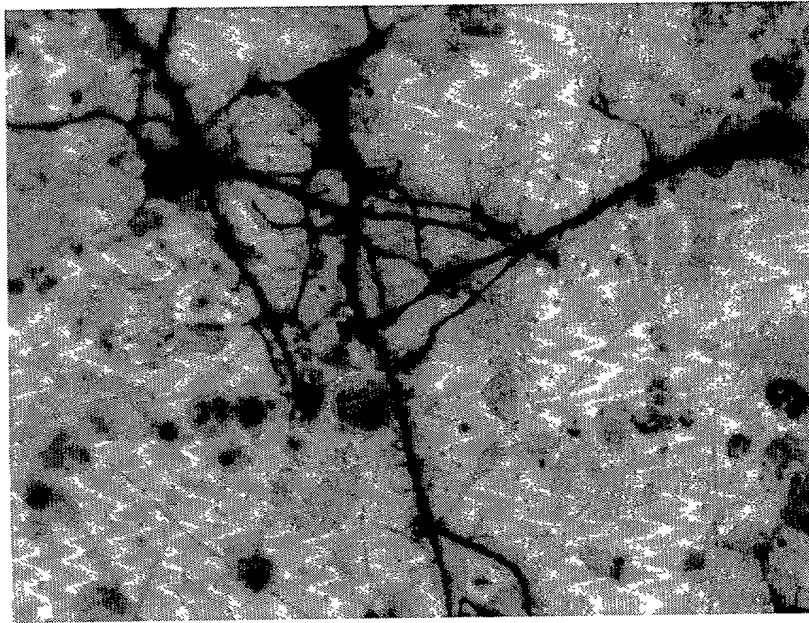
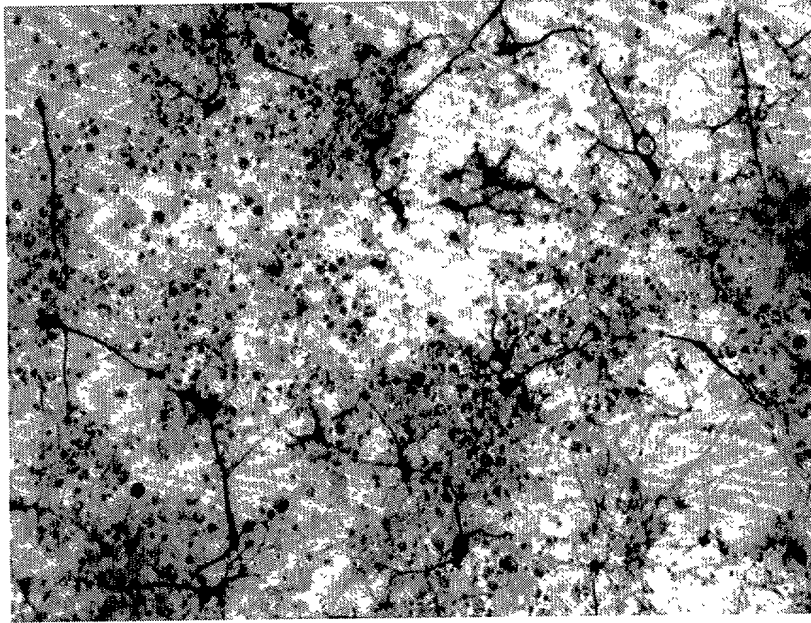


FIG. 3L

FIG. 4

Stability of Neuronal Differentiation

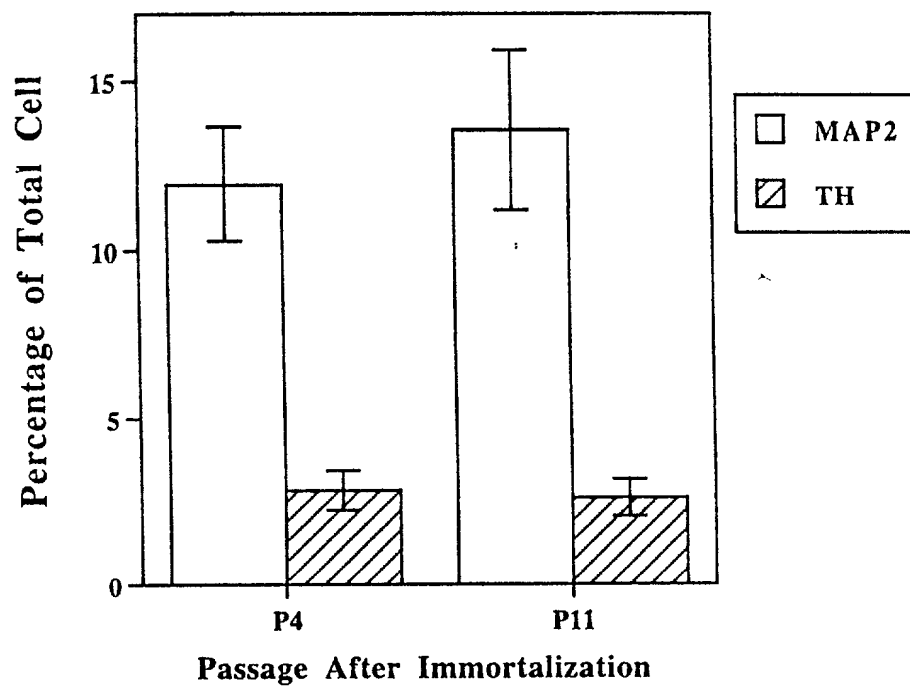


FIG. 5A

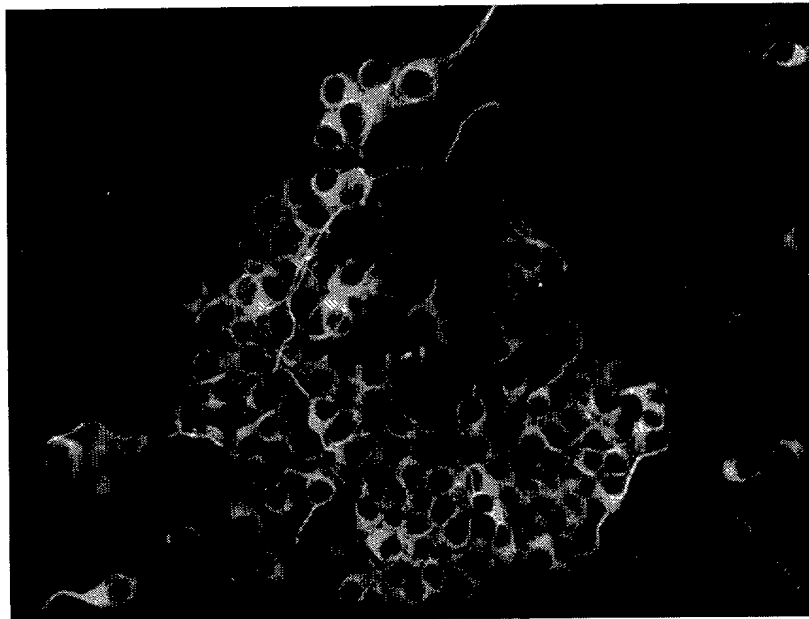
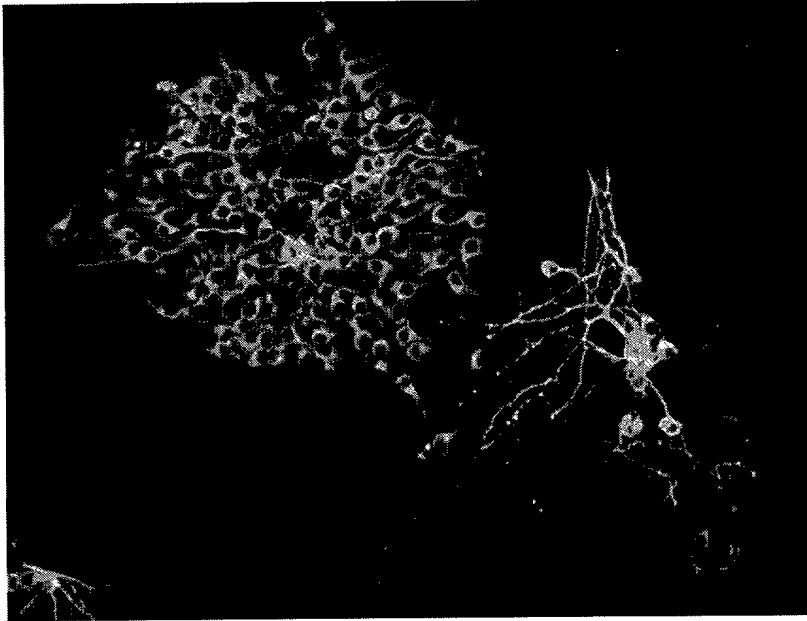


FIG. 5B

FIG. 5C

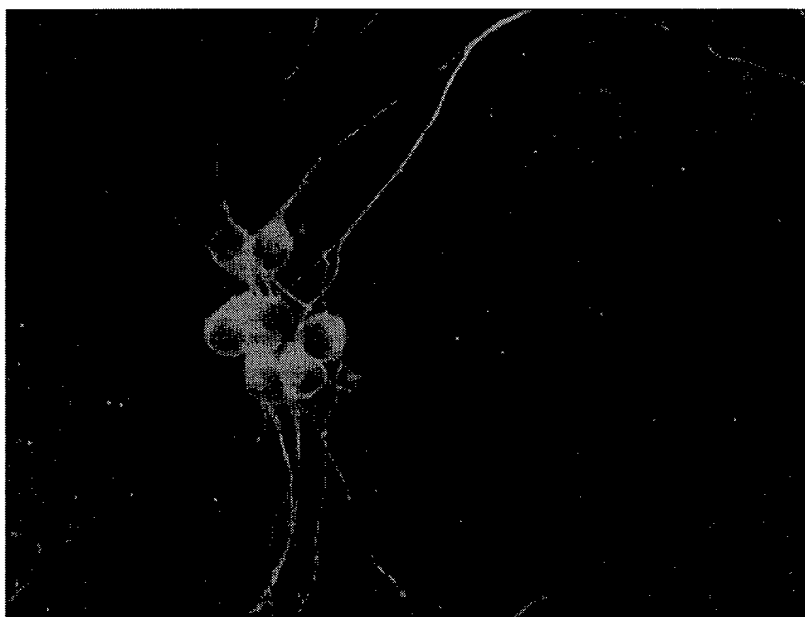
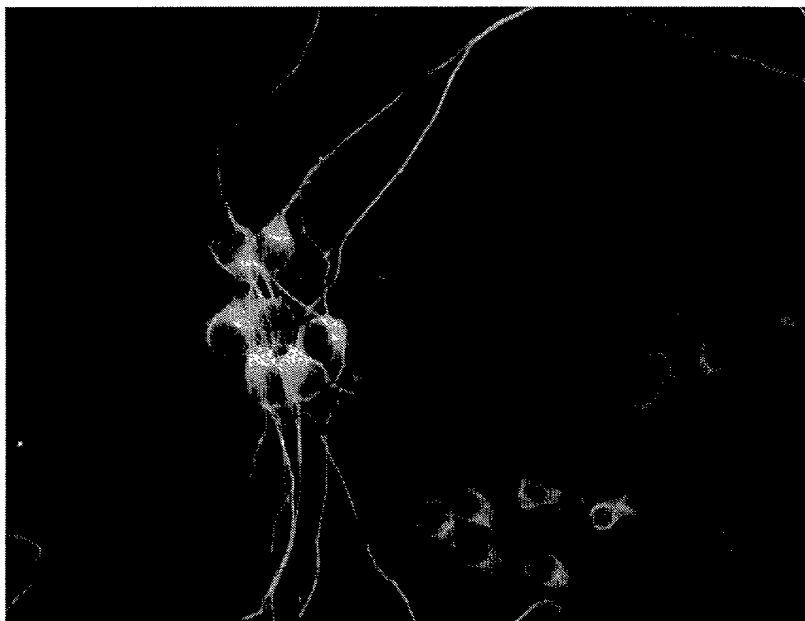


FIG. 5D

09398897, 09398897

Docket No.
0109015/016

Declaration and Power of Attorney For Patent Application

English Language Declaration

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name,

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

STABLE NEURAL STEM CELL LINES

the specification of which

(check one)

☒ is attached hereto.

☐ was filed on _____ as United States Application No. or PCT International Application Number _____ and was amended on _____ (if applicable)

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations, Section 1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, Section 119(a)-(d) or Section 365(b) of any foreign application(s) for patent or inventor's certificate, or Section 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate or PCT International application having a filing date before that of the application on which priority is claimed.

Prior Foreign Application(s)

Priority Not Claimed

_____ (Number)	_____ (Country)	_____ (Day/Month/Year Filed)	<input type="checkbox"/>
_____ (Number)	_____ (Country)	_____ (Day/Month/Year Filed)	<input type="checkbox"/>
_____ (Number)	_____ (Country)	_____ (Day/Month/Year Filed)	<input type="checkbox"/>

I hereby claim the benefit under 35 U.S.C. Section 119(e) of any United States provisional

60/101,354

(Application Serial No.)

September 22, 1998

(Filing Date)

(Application Serial No.)

(Filing Date)

(Application Serial No.)

(Filing Date)

I hereby claim the benefit under 35 U. S. C. Section 120 of any United States application(s), or Section 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. Section 112, I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, C. F. R., Section 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application:

09/053,414

(Application Serial No.)

April 1, 1998

(Filing Date)

pending

(Status)
(patented, pending, abandoned)

08/719,450

(Application Serial No.)

September 25, 1996

(Filing Date)

patented

(Status)
(patented, pending, abandoned)

(Application Serial No.)

(Filing Date)

(Status)
(patented, pending, abandoned)

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith. *(list name and registration number)*

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Patent and Trademark Office-U.S. DEPARTMENT OF COMMERCE

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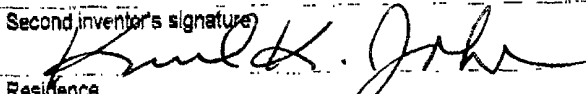
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